

BF₃ etherate-induced formation of C_{3–11}-alkenyl 2,3-unsaturated glucosides

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BF₃ etherate-induced formation of C_{3–11}-alkenyl 2,3-unsaturated glucosides was
used as the key step in their synthesis from glucose and C_{3–C₁₁}-alkenols.

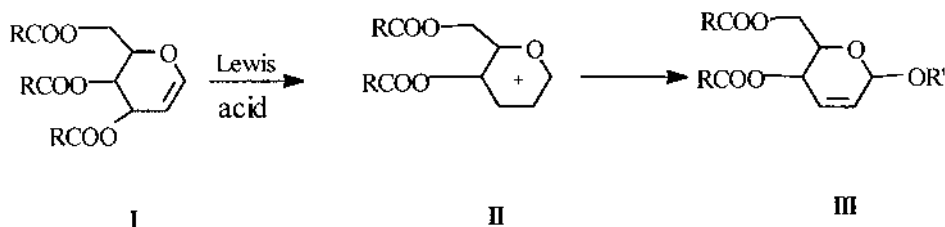
Keywords: synthesis of C_{3–11}-alkenyl 2,3-unsaturated glucosides, Ferrier reaction.

INTRODUCTION

Carbohydrates are involved in a large number of biological processes, *e.g.*, cell-recognition and regulatory processes.¹ In order to gain a better knowledge of the biological role of carbohydrates and their interactions with other biomolecules, *e.g.*, proteins, they have been extensively studied during the past decades. For many of these biological studies it was necessary to attach the carbohydrate to a carrier such as a protein or a lipid, *i.e.*, have a bifunctional spacer arm as a link between the carbohydrate moiety and the carrier in order to avoid shielding parts of the oligosaccharide structure from the carrier. Since the first synthetic glucoprotein was reported in 1929,^{3,4} many different spacer arms and methods for coupling oligosaccharides to different carrier have been developed.^{2,5,6} Micelles (liposomes) offer an alternative method for biological studies of carbohydrates. Furthermore, amphiphilic molecules such as long chain alkyl glucosides, can form a variety of different liquid crystalline phases some of which are believed to play a vital role in biological processes such as membrane fusion and endocytosis.^{7,8}

Since their implementation in 1988,⁹ *n*-pentenyl glucosides have proved valuable for the synthesis of oligosaccharides^{10,11} and enantiopure compounds¹² for mechanistic studies of glucosides hydrolysis^{9,13} and electrophilic additions,¹⁴ and for determining the relative reactivities of differently protected saccharides.¹⁵ The olefinic moi-

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Scheme 1.

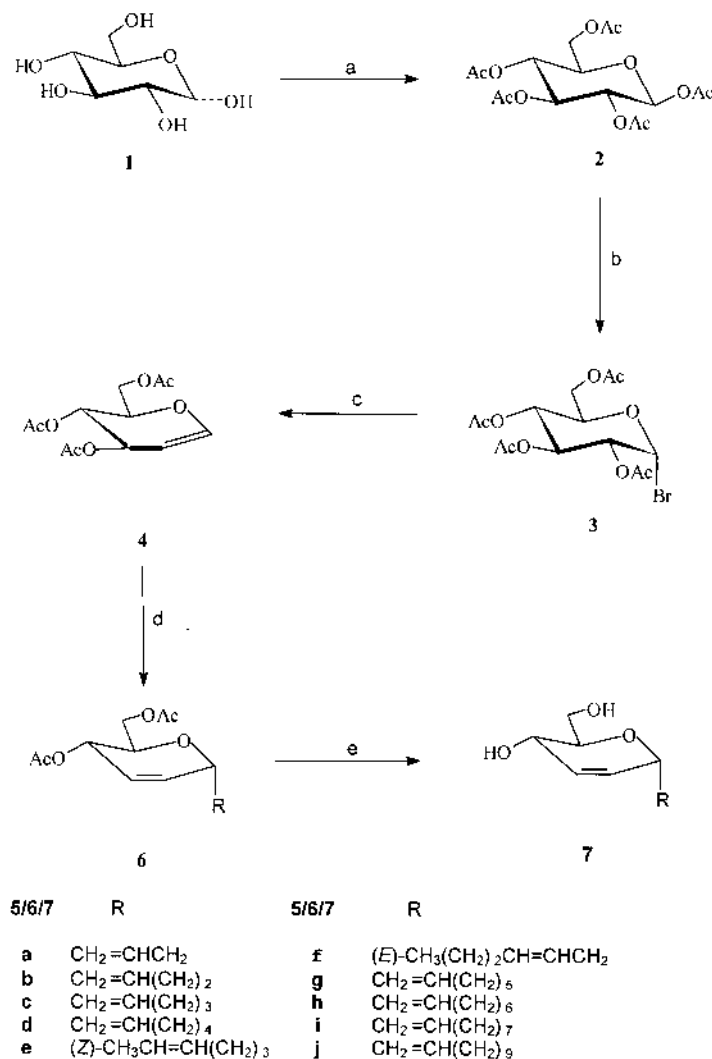
ety should allow for ready transformation into spacer functionalities, as has been exemplified by Vliegthart and co-workers.¹⁶ Versatility would, therefore, be enhanced, because after having served as a donor or acceptor in the synthesis of a saccharide, a given *n*-pentenyl glucoside could further serve for the elaboration of the spacer arm. With this in mind, we intended to synthesize some C₃–C₁₁-alkenyl 2,3-unsaturated glucosides starting from renewable material (glucose). The Lewis acid-catalyzed rearrangement of glycols in the presence of alcohols, known as the Ferrier reaction,¹⁷ is the method of choice for this synthesis. The reaction, as originally stated by Ferrier, involves an intermediate cyclic allylic oxocarbenium ion **II** (Scheme 1) to which the nucleophile adds preferentially in the quasi-axial orientation. The most commonly employed Lewis acid to effect this transformation is boron trifluoride etherate (BF₃ · Et₂O).

RESULTS AND DISCUSSION

In our previous work,¹⁸ it was demonstrated that some alkenols, under the influence of Lewis acids, such as BF₃·Et₂O, SnCl₄ and AlCl₃, cyclize to tetrahydrofuran and/or tetrahydropyran type ethers. For this reason it was considered interesting to investigate the competitiveness of the Ferrier reaction and cyclization in the case of alkenols. The present work has been shown that under the employed reaction conditions (short reaction times, low temperature), the Ferrier reaction prevails over the cyclization of the alkenols. As consequence of this, C₃–C₁₁-alkenyl 2,3-unsaturated glucosides have been successfully synthesized from glucose and C₃–C₁₁-alkenols using the Ferrier reaction. As depicted in the synthetic Scheme 2, treatment of glucose **1** with Ac₂O and molten anhydrous NaOAc at 120 °C gives exclusively the *-D*-glucose pentaacetate (**2**) which on treatment with HBr/AcOH leads to glucosyl bromide **3**. Reductive elimination with Zn-Cu affords tri-*O*-acetyl glucal (**4**). Glycosylation with alkenol **5** and boron trifluoride etherate proceeds by nucleophilic attack at the anomeric center of the glucal **4**. By an allylic rearrangement, the activated ester function at C-3 leaves the molecule to give the alkenyl 2,3-unsaturated glucoside **6** / with the *-*anomer largely prevailing (**6** : **6** = 7 : 1).

Following chromatographic purification the peracetylated products are deacetylated with the mixture methanol : triethylamine : water (2 : 1 : 1) to give alkenyl *-*glucoside (**7**).

For structural assignments extensively NMR studies were employed (¹H -¹H homonuclear and ¹H -¹³C heteronuclear chemical correlation spectroscopy (COSY) experiments were performed).



a) Ac₂O, NaOAc b) HBr, AcOH c) Cu-Zn d) ROH (**3**) e) MeOH-Et₃N-H₂O 2:1:1

Scheme 2

As starting alcohols some primary alkenols were used. For this reason, 3-buten-1-ol (**5b**), 4-penten-1-ol (**5c**), 5-hexen-1-ol (**5d**), 6-hepten-1-ol (**5g**), 7-octen-1-ol (**5h**) and 8-nonen-1-ol (**5i**) were synthesized according to known procedures.

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EXPERIMENTAL

General

Column chromatography: Merck silica gel 60, particle size 0.04–0.06 mm. *IR-spectra:* Perkin Elmer Model 137B spectrophotometer. *NMR spectra:* Varian Gemini 200 (200 MHz) in CDCl₃ or Bruker 250 (62.9 MHz) in DMSO-d₆. *Mass spectra:* finnigan Mat 8320 spectrometer.

Reactants and products

The substrate alcohol 4-penten-1-ol (**5c**), b.p. 135–136 °C, was prepared from tetrahydrofurfuryl alcohol, *via* tetrahydrofurfuryl chloride.¹⁹ 5-Hexen-1-ol (**5d**) (b.p. 63 °C at 17 mbar) was prepared in several steps in 15 % overall yield: the Grignard reaction of allyl chloride with magnesium and paraformaldehyde gave 3-buten-1-ol (**5b**, 46 %),^{20,21} which was converted by means of phosphorous tribromide to 1-bromo-3-butene (73 %).²⁰ Alkylation of diethyl malonate with this 1-bromo-3-butene (70 %) and decarboxylation of the formed diester by the DMSO-NaCl general procedure (76 %),²² followed by reduction of the resulting ethyl ester of 5-hexenoic acid with LiAlH₄ in diethyl ether (93 %), 5-hexen-1-ol (**5d**) was obtained. 6-Hepten-1-ol (**5g**), 7-octen-1-ol (**5h**) and 8-nonen-1-ol (**5i**) were prepared from the corresponding dicarboxylic acids (suberic, azelaic and sebacic acid, respectively): decarboxylation of their mono-methyl esters²³ by means of lead tetraacetate.²⁴ Lead tetraacetate (LTA) gave the methyl esters of the terminally unsaturated monocarboxylic acids, which were reduced with LiAlH₄ in diethyl ether. The tri-*O*-acetyl glucal (**4**) was prepared according to the literature procedure,^{25,26} *via* acetylated sugar **2** and glucosyl bromide **3** (Scheme 2).

Prop-2-enyl 4,6-di-O-acetyl-2,3-dideoxy-α-D-erythro-hex-2-enopyranoside (6a)

General procedure. To a stirred solution of glucal (**4**, 1.36 g, 5.00 mmol) in dry toluene (25 ml) were added 2-propen-1-ol (**5a**, 0.33 ml, 4.95 mmol) and a catalytic amount of BF₃ · Et₂O (0.4 ml, 3.25 mmol). The mixture was allowed to react for 1 h and then neutralized by addition of Na₂CO₃ (2 g). After the solution had been stirred for 30 min, the solids were filtered off and the filtrate was successively washed with a saturated aqueous solution of NaHCO₃ and distilled water. After drying with anhydrous Na₂SO₄, the solvent was evaporated on a rotary evaporator under vacuum affording a syrup (1.30 g). The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6a** in 96 % yield as a 7:1 mixture of *-* and *-*prop-2-en-1-yl glucosides. MS: *m/z* = 271 (M+H)⁺, 213, 153.

But-3-enyl 4,6-di-O-acetyl-2,3-dideoxy-α-D-erythro-hex-2-enopyranoside (6b)

Tri-*O*-acetyl glucal (**4**, 1.36 g, 5.00 mmol) in dry toluene (25 ml) was treated with 3-buten-1-ol (**5a**, 0.42 ml, 4.88 mmol) and a catalytic amount of BF₃ · Et₂O (0.4 ml, 3.25 mmol) as described for **6a**. The described work-up and removal of the solvent gave 1.15 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6b** in 83 % yield as a 7:1 mixture of *-* and *-*but-3-en-1-yl glucosides. MS: *m/z* = 285 (M+H)⁺, 213, 153.

Pent-4-enyl 4,6-di-O-acetyl-2,3-dideoxy-α-D-erythro-hex-2-enopyranoside (6c)

Tri-*O*-acetyl glucal (**4**, 1.36 g, 5.00 mmol) in dry toluene (25 ml) was treated with 4-penten-1-ol (**5c**, 0.5 ml, 4.84 mmol) and a catalytic amount of BF₃ · Et₂O (0.4 ml, 3.25 mmol) as described for **6a**. The described work-up and removal of solvent gave 1.25 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6c** in 87 % yield as a 6:1 mixture of *-* and *-*pent-4-en-1-yl glucosides. MS: *m/z* = 299 (M+H)⁺, 239 (M-60)⁺, 213, 153.

Hex-5-enyl 4,6-di-O-acetyl-2,3-dideoxy-α-D-erythro-hex-2-enopyranoside (6d)

Tri-*O*-acetyl glucal (**4**, 1.36 g, 5.0 mmol) in dry toluene (25 ml) was treated with 5-hexen-1-ol (**5d**, 0.6 ml, 5.08 mmol) and a catalytic amount of BF₃ · Et₂O (0.40 ml, 3.25 mmol) as described for **6a**. The described work-up and removal of the solvent gave 1.34 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6d** in 86 % yield as a 7:1 mixture of *-* and *-*hex-4-en-1-yl glucosides. MS: *m/z* = 313 (M+H)⁺, 253 (M+H-60)⁺, 213, 153.

Hex-(E)-2-enyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (6e).

Tri-*O*-acetyl glucal (**4**, 1.36 g, 5.0 mmol) in dry toluene (25 ml) was treated with (*E*)-2-hexen-1-ol (**5e**, 0.6 ml, 5.0 mmol) and a catalytic amount of BF₃ · Et₂O (0.40 ml, 3.25 mmol) as described for **6a**. The described work-up and removal of the solvent gave 1.27 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6e** in 81 % yield as a 7:1 mixture of - and -hex-(*E*)-2-en-1-yl glucosides. MS: m/z = 313 (M+H)⁺, 253 (M+H-60)⁺, 213, 153.

Hex-(Z)-4-enyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (6f).

Tri-*O*-acetyl glucal (**4**, 1.36 g, 5.0 mmol) in dry toluene (25 ml) was treated with (*Z*)-4-hexen-1-ol (**5f**, 0.6 ml, 5.0 mmol) and a catalytic amount of BF₃ · Et₂O (0.40 ml, 3.25 mmol) as described for **6a**. The described work-up and removal of the solvent gave 1.29 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6f** in 83 % yield as a 7:1 mixture of - and -hex-(*Z*)-4-en-1-yl glucosides. MS: m/z = 313 (M+H)⁺, 253 (M+H-60)⁺, 213, 153.

Hept-6-enyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (6g).

Tri-*O*-acetyl glucal (**4**, 0.680 g, 2.5 mmol) in dry toluene (25 ml) was treated with 6-hepten-1-ol (**5g**, 0.285 ml, 2.5 mmol) and a catalytic amount of BF₃ · Et₂O (0.20 ml, 1.60 mmol) as described for **6a**. The described work-up and removal of the solvent gave 0.81 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6g** in 99 % yield as a 9:1 mixture of - and -hept-6-en-1-yl glucosides. MS: m/z = 327 (M+H)⁺, 267 (M+H-60)⁺, 213, 153.

Oct-7-enyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (6h).

Tri-*O*-acetyl glucal (**4**, 1.36 g, 5.0 mmol) in dry toluene (25 ml) was treated with 7-octen-1-ol (**5h**, 0.640 ml, 4.99 mmol) and a catalytic amount of BF₃ · Et₂O (0.4 ml, 3.25 mmol) as described for **6a**. The described work-up and removal of the solvent gave 1.41 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6h** in 83 % yield as a 7:1 mixture of - and -oct-7-en-1-yl glucosides. MS: m/z = 314 (M+H)⁺, 281 (M+H-60)⁺, 213, 153.

Non-8-enyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (6i).

Tri-*O*-acetyl glucal (**4**, 0.680 g, 2.5 mmol) in dry toluene (12.5 ml) was treated with 8-nonen-1-ol (**5i**, 0.360 ml, 2.59 mmol) and a catalytic amount of BF₃ · Et₂O (0.20 ml, 1.62 mmol) as described for **6a**. The described work-up and removal of the solvent gave 0.850 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6i** in 96 % yield as a 6:1 mixture of - and -non-8-en-1-yl glucosides. MS: m/z = 355 (M+H)⁺, 295 (M+H-60)⁺, 213, 153.

Undec-10-enyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (6j).

Tri-*O*-acetyl glucal (**4**, 1.35 g, 5.0 mmol) in dry toluene (12.5 ml) was treated with 10-undecen-1-ol (**5j**, 1.0 ml, 5.0 mmol) and a catalytic amount of BF₃ · Et₂O (0.40 ml, 3.25 mmol) as described for **6a**. The described work-up and removal of the solvent gave 1.43 g of a syrupy residue. The residue was purified by dry flash chromatography (toluene–ethyl acetate) to afford **6j** in 85 % yield as a 6:1 mixture of - and -undec-8-en-1-yl glucosides. MS: m/z = 383 (M+H)⁺, 323 (M+H-60)⁺, 213, 153.

The resulting material was deacetylated by treatment of the purified sample (100–200 mg) with 2 ml of MeOH–Et₃N–H₂O 2:1:1 at room temperature for 24 hours to give alkenyl 2,3-dideoxy- α -D-erythro-hex-2-enopyranosides (**7**).

TABLE II. ¹³C-NMR data of compounds **6a-j** (50.0 MHz, CDCl₃) and **7a, b, e** (62.9 MHz, DMSO-d₆).

Carbon	6a	6b-d, 6g-j	6e	6f	7a	7b	7e
C-1	93.4	94.1–94.4	92.9	94.3	93.4	93.5	92.8
C-2	129.0	128.7–129.0	128.8	128.8	135.6	135.8	134.7
C-3	127.6	127.7–127.8	127.7	127.8	134.8	134.4	133.5
C-4	65.1	65.1–65.2	65.0	65.1	73.1	72.8	72.9
C-5	66.7	66.6–66.8	66.6	66.7	62.4	62.4	62.6
C-6	62.7	62.8–63.0	62.7	62.9	61.4	61.0	61.1
C-1'	69.0	68.0–68.8	68.7	68.2	68.2	66.8	67.9
(CH ₂) _n		25.4–34.0	21.8, 34.1	23.2, 29.4		33.8	22.1, 34.0
CH=CH ₂	117.2	113.9–116.5			117.2	116.4	
CH=CH			125.4, 135.1	124.4, 129.5			125.6, 126.9
CH=CH ₂	133.9	134.9–138.9			126.0	125.5	
CHCH ₃			13.4	12.6			13.7

ИЗВОД

BF₃-ЕТЕРАТОМ ИНДУКОВАНО ФОРМИРАЊЕ C₃-C₁₁-АЛКЕНИЛ-2,3-НЕЗАСИЋЕНИХ ГЛУКОЗИДАСТАНИМИР КОНСТАНТИНОВИЋ^а, ЈАСМИНА ПРЕДОЈЕВИЋ^а, СВЕТИСЛАВ ГОЈКОВИЋ^б и
ВЛАДИМИР ПАВЛОВИЋ^в^аИнститут за хемију Природно-мајематичког факултета Универзитета у Крагујевцу, п. бр. 60, 34000 Крагујевац, E-mail: konstan@eunet.yu, ^бИнститут за хемију, технологију и металургију, Ђежосева 12, 11000 Београд, ^вХемијски факултет Универзитета у Београду, Сидунски прџ 16, п. бр. 158, 11001 БеоградBF₃-Етератом индуковано формирање C₃-C₁₁-алкенил-2,3-незасићених глюкозида је примењено у њиховој вишестепеној синтези која полази од глюкозе и C₃-C₁₁-алкенола.

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REFERENCES

1. A. Varki, *Glucobiology* **3** (1993) 97
2. E. V. Bovin, H. -J. Gabius, *Chem. Soc. Rev.* (1995) 413
3. W. F. Goebel, O. T. Avery, *J. Exp. Med.* (1929) 521
4. O. T. Avery, W. F. Goebel, *J. Exp. Med.* (1929) 533
5. C. P. Stowel, Y. C. Lee, *Adv. Carbohydr. Biochem.* **37** (1980) 225
6. J. D. Aplin, J. C. Wriston, *Crit. Rev. Biochem.* (1981) 259
7. R. E. Marchant, M. Rueggsegger, Y. Qui, in *Polysaccharides*, S. Dumitriu, Ed., Dekker, New York, 1998, p. 851
8. J. -H. Fuhrhop, B. Rosengarten, *Synth. Lett.* (1997) 1015
9. D. R. Mootoo, V. Date, B. Fraser-Reid, *J. Am. Chem. Soc.* **110** (1988) 2662
10. B. Fraser-Reid, U. E. Udodong, Z. Wu, H. Ottoson, J. R. Merrit, C. S. Rao, C. Roberts, R. Madsen, *Synth. Lett.* (1992) 927
11. R. Madsen, B. Fraser-Reid, in *Modern Methods in Carbohydrate Chemistry*, Harwood Academic Publishers, Switzerland, 1995

12. J. M. Llera, C. J. Lopes, B. Fraser-Reid, *J. Org. Chem.* **55** (1990) 2997
13. R. Rodebaugh, B. Fraser-Reid, *Tetrahedron* **52** (1996) 7663
14. B. Fraser-Reid, Z. Wu, U. E. Udodong, H. Ottoson, *J. Org. Chem.* **55** (1990) 6068
15. d. r. Mootoo, P. Konradsson, U. E. Udodong, B. Fraser-Reid, *J. Am. Chem. Soc.* **110** (1988) 5583
16. P. B. van Seeventer, J. A. L. M. van Dorst, J. F. Semerink, J. P. Kamerling, J. F. G. Vliegthart, *Carbohydr. Res.* **300** (1997) 369
17. R. J. Ferrier, *Adv. Carbohydr. Chem. Biochem.* **24** (1996) 199, and references cited therein
18. M. Lj. Mihailović, Z. Petrović, A. Teodorović, S. Konstantinović, V. Andrejević, *C. R. Acad. Sci. Paris, Serie II* **308** (1989) 29
19. L. A. Brooks, H. R. Snyder, *Org. syntheses*, Coll. Vol. **3** (1955) 698
20. R. P. Linstead, H. N. Rydon, *J. Chem. Soc.* (1934) 1998
21. A. Amtstutz, *J. Org. Chem.* **9** (1944) 310
22. A. P. Krapcho, a. J. Lovey, *Tetrahedron Lett.* (1973) 957
23. L. J. Durham, D. J. McLeod, J. Casan, *Org. Synthesis*, Coll. Vol **4** (1963) 635
24. J. D. Bacha, J. K. Kochi, *Tetrahedron* **24** (1968) 2215; Yu. N. Ogibin, M. I. Katzin, G. I. Nikishin, *Synthesis* (1974) 889
25. B. Iselin, T. Reichstein, *Helv. Chim. Acta* **27** (1944) 1146, 1200
26. W. Roth, W. Pigman, *Methods in Carbohydr. Chem.* **2** (1963) 405.

TABLE I. ¹H-NMR data of the compounds **6a-j** (200 MHz, CDCl₃)^{a)}

	6a	6b	6c	6d	6e	6f	6g	6h	6i	6j
H-1. CH=CH ₂	5.06–5.38	4.98–5.33	4.93–5.10	4.92–5.12		5.28–5.53	4.90–5.07	4.83–5.08	4.88–5.12	4.90–5.10
H-2,3; CH=CH ₂	5.7–6.0	5.70–5.94	5.72–5.96	5.71–5.97	5.51–5.97	5.80–6.0	5.70–5.96	5.64–5.92	5.70–5.97	5.70–5.94
H-4	5.06–5.38	5.33	5.32	5.31	5.31		5.31	5.27	5.32	5.31
H-5,6	4.0–4.32	4.03–4.26	4.0–4.3	4.0–4.3	3.96–4.32	4.0–4.31	4.0–4.3	3.97–4.25	4.01–4.31	4.0–4.3
CH=CH					5.51–5.97	5.28–5.53				
H-1a'	4.0–4.32	3.56	3.53	3.52		3.51	3.50	3.46	3.51	3.54
H-1b'	4.0–4.32	3.81	3.80	3.78		3.79	3.78	3.72	3.77	3.78
H-2'		2.32	1.72	1.62		1.69	1.61	1.57	1.58	1.58
CH ₂ CH=C			2.12	2.08		2.0–2.2	2.06	2.05	2.07	2.02
CH ₃ CO	2.08, 2.08	2.06, 2.07	2.09, 2.10	2.09, 2.10	2.08, 2.10	2.09, 2.10	2.08, 2.10	2.04, 2.05	2.09, 2.10	2.07, 2.09
(CH ₂) _n				1.50	1.41		1.40	1.30	1.33	1.30
CH ₃ CH					0.91	1.61				

^{a)} In all compounds the coupling constants in the 2,3-unsaturated glucosyl ring are approximately the same as follows: ³J_{3,4} = 0.8 Hz, ³J_{4,5} = 9.6 Hz, ³J_{5,6a} = 2.4 Hz, ³J_{5,6b} = 5.1 Hz, ²J_{6a,6b} = -11.7 Hz, ³J_{1a',2'} = ³J_{1b',2'} = 6.3 Hz, ²J_{1a',1b'} = 9.6 Hz.